Quick Start Guide

GenElute™ Gel Extraction Kit

NA1111

Reagents to Prepare

- Dilute the Wash Solution Concentrate G with 48 mL of 95-100% ethanol. After each use, tightly cap the diluted Wash Solution to prevent ethanol evaporation.

Protocol

All spins at 12,000-16,000 x g.

Sample Preparation

1. Excise the DNA fragment of interest and weigh the gel slice in a colorless tube.
2. Add 3 gel volumes of Gel Solubilization Solution and incubate the mixture at 50-60 °C for 10 min or until the gel slice is completely dissolved. Vortex every 2-3 min during incubation to dissolve the gel.
3. Once gel slice is dissolved ensure the color of the mixture is yellow. If the mixture color is red, add 10 µL of 3M Sodium Acetate, pH 5.2 and mix. Continue adding 3M Sodium Acetate, pH 5.2 in increments of 10 µL until the mixture turns yellow.
4. Add 1 gel volume of 100% isopropanol and mix until homogenous. If the agarose concentration is > 2%, use 2 gel volumes of 100% isopropanol.

Bind DNA

5. Place Column into a 2 mL collection tube.
6. Add 0.5 mL Column Preparation Solution to each column and spin for 1 min. Discard the flowthrough.
7. Load 700 µL solubilized gel mixture into column and spin for 1 min. Discard flowthrough. Repeat if the volume is > 700 µL.

Wash to Remove Contaminants

8. Add 700 µL of Wash Solution to the column and spin for 1 min. Discard the eluate.
9. Replace column with its collection tube and spin for an additional 1 min to remove excess ethanol. Discard the eluate and collection tube.

DNA Elution

10. Transfer column to fresh collection tube.
11. Add 50 µL Elution Solution (pre-heated to 65 °C) to the center of column. Incubate at room temperature for 1 min.
12. Spin column for 1 min.

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