

SIGMA QUALITY CONTROL TEST PROCEDURE

ProductInformation

Enzymatic Assay of ASPARAGINASE (EC 3.5.1.1)

PRINCIPLE:

L-Asparagine + H₂O Asparaginase > L-Aspartate + NH₃

CONDITIONS: $T = 37^{\circ}C$, pH = 8.6, A_{436nm} , Light path = 1 cm

METHOD: Spectrophotometric Stop Rate Determination

REAGENTS:

A. 50 mM Tris Buffer, pH 8.6 at 37°C (Prepare 100 ml in deionized water using Trizma Base, Prod. No. T-1503. Adjust to pH 8.6 at 37°C with 1 M HCI.)

- B. 189 mM L-Asparagine Solution (Prepare 10 ml in deionized water using L-Asparagine, Anhydrous, Prod. No. A-0884.)
- C. 6 mM Ammonium Sulfate Standard Solution ((NH₄)₂SO₄ Std) (Prepare 100 ml deionized water using Ammonium Sulfate, Grade I, Prod. No. A-5132.)
- D. 1.5 M Trichloroacetic Acid (TCA) (Prepare 10 ml in deionized water using Trichloroacetic Acid, 6.1 N Solution, Stock No. 490-10.)
- E. Ammonia Color Reagent (Use Nessler's Reagent, Aldrich Stock No. 34,518-8.)
- F. Asparaginase Enzyme Solution (Immediately before use, prepare a solution containing 2.0 4.0 units/ml of Asparaginase in cold deionized water.)

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PROCEDURES:

Step 1:

Pipette (in milliliters) the following reagents into suitable tubes:

	Test	Blank_	<u>Std 1</u>	<u>Std 2</u>	Std 3	Std <u>Blank</u>					
Reagent A (Buffer) Reagent B (L-Asparagine Soln) Reagent C	1.00	1.00	1.00	1.00	1.00	1.00					
	0.10	0.10									
$((NH_4)_2SO_4Std)$			0.25	0.50	1.00						
Deionized Water	0.90	0.90	0.85	0.60	0.10	1.10					
Equilibrate to 37°C. Then add:											
Reagent F (Enzyme Solution)	0.10										
Immediately mix by inversion and incubate at 37°C for 30 minutes. Then add:											
Reagent D (TCA) Reagent F	0.10	0.10	0.10	0.10	0.10	0.10					
(Enzyme Solution)		0.10									

Mix by inversion. Centrifuge for 2 minutes to clarify.

Step 2:

Pipette (in milliliters) the following reagents into suitable containers:

Deionized Water Supernatant	4.30	4.30	4.30	4.30	4.30	4.30
(from Step 1)	0.20	0.20	0.20	0.20	0.20	0.20
Reagent E (Ammonia Color Reagent)	0.50	0.50	0.50	0.50	0.50	0.50

Immediately mix by inversion and after 1 minute record the A_{436nm} for Standards, Tests, and Blanks.

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CALCULATIONS:

Standard Curve:

 ΔA_{436nm} Standard = A_{436nm} Standard - A_{436nm} Standard Blank

Prepare a standard curve by plotting the ΔA_{436nm} of the Standard versus Ammonia (NH₃) concentration. Note that 1 mole of Ammonium Sulfate corresponds to 2 moles of Ammonia, therefore a 6 mM Ammonium Sulfate standard is equivalent to a 12 mM ammonium standard.

Sample Determination:

$$\Delta A_{436nm}$$
 Test = A_{436nm} Test - A_{436nm} Test Blank

Determine the µmoles of NH₃ liberated using the standard curve.

Units/ml enzyme =
$$\frac{(\mu \text{mole of NH}_3 \text{ liberated})(2.20)}{(0.2)(30)(0.1)}$$

2.20 = Volume of Step 1

0.2 = Volume of Step 1 used in Step 2

30 = Time of assay in minutes

0.1 = Volume (in milliliters) of enzyme used

UNIT DEFINITION:

One unit will liberate 1.0 µmole of ammonia from L-asparagine per minute at pH 8.6 at 37°C.

FINAL ASSAY CONCENTRATION:

In a 2.20 ml reaction mix, the final concentrations are, 23 mM Tris, 8.6 mM L-asparagine and 0.2 - 0.4 units of asparaginase.

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REFERENCES:

Shirfrin, S., Parrott, C.L. and Luborsky, S.W. (1974) Journal of Biological Chemistry 249, 1335-1340

NOTES:

- 1. This assay is based on the cited reference.
- Where Sigma Product or Stock numbers are specified, equivalent reagents may be substituted.

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